

A comparative analysis of mono- and disaccharide benzyl fucopyranosides

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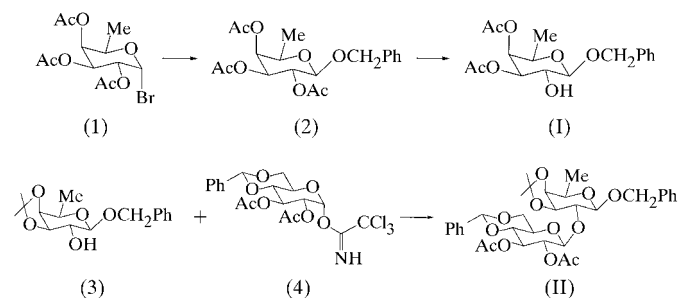
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The syntheses and X-ray analyses of two fucopyranosides, the monosaccharide benzyl 3,4-di-*O*-acetyl-2-hydroxy- β -D-fucopyranoside, C₁₇H₂₂O₇, and the disaccharide 1-benzyl *O*-(2,3-di-*O*-acetyl-4,6-*O*-benzylidene- β -D-glucopyranosyl)-(1 \rightarrow 2)-3,4-*O*-isopropylidene- β -D-fucopyranoside, C₃₃H₄₀O₁₂, are described. The different substituents induce small conformational changes on the fucopyranoside ring. However, the conformation of the benzyl group varies from (+)*gauche* for the monosaccharide to synperiplanar for the disaccharide.

Comment

Fucose is an important deoxysugar found recently as a constituent unit of glycoresins from the genus *Ipomea* (Bah & Pereda-Miranda, 1997). These glycoresins, called tricolorins, have attracted much interest from biological and chemical points of view. As part of a strategy directed towards the synthesis of analogues of the corresponding glycosides, the syntheses and X-ray analyses of benzyl fucopyranoside, (I) (Fig. 1), and the closely related benzyl disaccharide, (II) (Fig. 2), have been performed and the results are presented here.



In both compounds, the pyranoside rings have a ⁴C₁ chair conformation, with all substituents oriented in equatorial positions, except for the substituent at position C4 of the glucopyranoside moiety, which exhibits an axial orientation. For compound (II), the equatorial orientation at position 3 in

the fucopyranoside ring is frustrated by the formation of a five-membered ring. Therefore, a biaxial conformation is observed, with an angle of 53.9 (2)° between the C3—O3 bond and the Cremer–Pople plane, calculated according to Cremer & Pople (1975) and defined, in both compounds, by the atoms C1–C5 and O5.

The isopropylidene group also produces the significant deviation from the ideal ⁴C₁ chair conformation observed in the fucopyranoside ring of compound (II), as can be seen when comparing the puckering parameters (Cremer & Pople, 1975) of both fucopyranoside rings [$Q = 0.572$ (2) Å, $\theta = 176.9$ (2)° and $\varphi = 186$ (3)° for (I), and $Q = 0.556$ (3) Å, $\theta = 159.9$ (3)° and $\varphi = 220.9$ (8)° for (II)]. In compound (II), the effect of the benzylidene group on the glucopyranosil

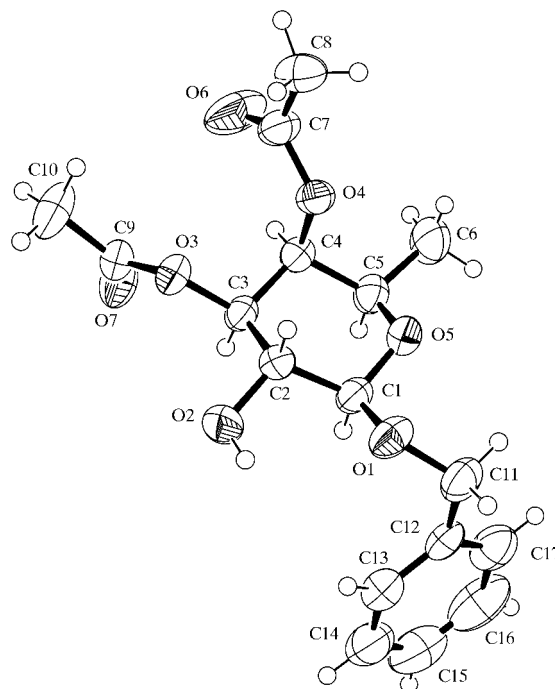


Figure 1

A view of the molecular structure of (I), showing 50% displacement ellipsoids and the atom-numbering scheme. H atoms are drawn as small spheres of arbitrary radii.

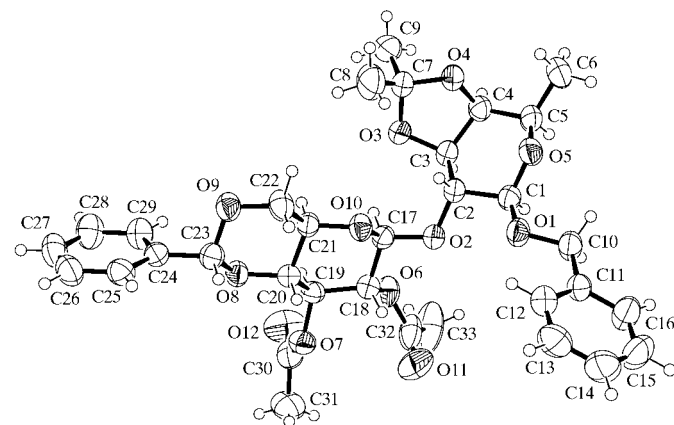


Figure 2

A view of the molecular structure of (II), showing 50% displacement ellipsoids and the atom-numbering scheme. H atoms are drawn as small spheres of arbitrary radii.

conformation is smaller, and the ring defined by atoms O10 and C17–C21 is very close to the ideal 4C_1 chair conformation [$Q = 0.597$ (3) Å, $\theta = 170.7$ (3)° and $\varphi = 156.0$ (16)°].

The angle between the glycosidic bond and the normal to the Cremer–Pople plane, calculated for both fucopyranoside rings, is 68.7 (1)° for (I) and 76.0 (2)° for (II). Due to the exoanomeric effect, the substituent on C1, in this case the benzyl group, should prefer a (+)gauche conformation (Jeffrey, 1990). This is the case in (I), as can be seen in the C12–C11–O1–C1 torsion angle, but not in (II), which shows a clear synperiplanar conformation, with atoms C1, O1, C10 and C11 in almost the same plane [largest deviation from the least-squares plane is 0.023 (1) Å for atom O1]. It is also noticeable that, for (I), the C5–O5, C1–O5 and C1–O1 bond distances (Table 1) are closer to the standard values for an axial glycosidic bond (1.434, 1.419 and 1.398 Å, respectively) than for an equatorial one (1.426, 1.428 and 1.385 Å, respectively) (Jeffrey & Taylor, 1980; Jeffrey, 1990). In (II), these bond distances (Table 3) do not correspond clearly to equatorial or axial cases.

In the disaccharide, (II), atoms O5, C1, C2, O2, C17, C18 and C19 are almost coplanar [largest deviation from the least squares plane is –0.109 (2) Å for C19]. Therefore, three atoms of each saccharide lie in the plane defined by the glycosidic linkage. For each pyranoside, the rest of the atoms lie on one side of this plane, the two rings being on opposite sides of the plane. For this reason, the O10–C17–O2–C2 torsion angle is almost equal to C3–C2–O2–C17. This conformation could be partly due to the weak hydrogen bond joining C17 in the glucopyranoside ring and O3 in the fucopyranoside moiety (Table 4).

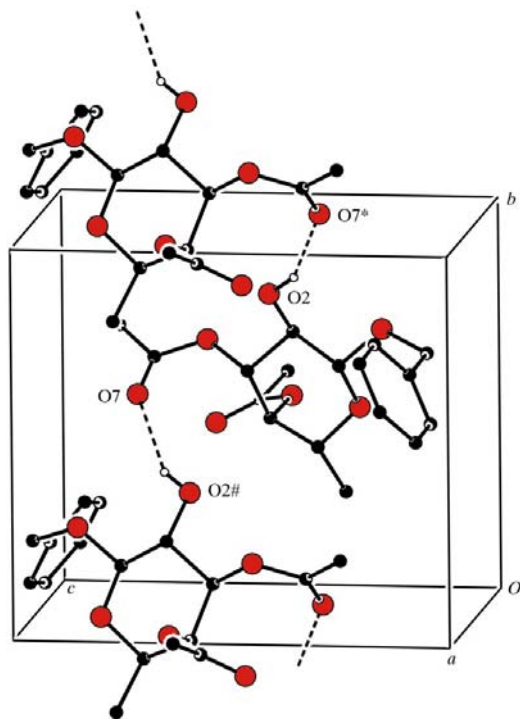


Figure 3
A drawing of the unit cell of (I), viewed along the *a* axis. Dashed lines indicate hydrogen bonds. Atom O7* is at symmetry position $(2 - x, y + \frac{1}{2}, 1 - z)$ and atom O2# is at symmetry position $(2 - x, y - \frac{1}{2}, 1 - z)$.

An O–H···O hydrogen bond is observed in (I) between the hydroxy group and one of the acetate O atoms (Table 2). Due to this intermolecular interaction, the molecules form chains along the *b* direction (Fig. 3). There are also weak C–H···O interactions involving the benzyl group as donor and the pyranoside group as acceptor, which join the chains along the *a* direction. For compound (II), there are no strong hydrogen-bond interactions, due to the absence of hydroxyl groups. The only significant intermolecular interaction is a weak C–H···O hydrogen bond (Table 4) joining the molecules in the *a* direction.

Experimental

Compound (I) (see *Scheme*) was prepared using fucose (Lerner, 1993) as the starting material, which was peracetylated with acetic anhydride. Bromination of the anomeric C atom was performed using hydrobromic acid (30 wt%) in acetic acid to produce intermediate (1), which was subsequently reacted with benzyl alcohol under Koenigs–Knorr conditions to yield glycoside (2) (Igarashi, 1977). Deacetylation with sodium methoxide solution afforded a compound which was treated with two equivalents of acetic anhydride to produce (I). Spectroscopic analysis for (I), ${}^1\text{H}$ NMR (CDCl_3 , 300 MHz, δ , p.p.m.): 1.3 (*d*, 3H), 2.0 and 2.2 (both *s*, 3H), 3.8 (*m*, 1H), 3.9 (*dd*, 1H), 4.4 (*d*, 1H), 4.6 (*d*, 1H), 4.95 (*dd*, 1H), 5.0 (*d*, 1H), 5.22 (*dd*, 1H), 7.4 (*m*, 5H). Compound (II) (see *Scheme*) was prepared under Fischer *O*-glycoside coupling-reaction conditions, by the reaction between fucose and benzyl alcohol under a controlled stream of HCl gas to produce benzyl fucose (Searle *et al.*, 1996). Acetonide formation was performed under standard conditions (acetone and 2,2-dimethoxypropane, with *p*-toluenesulfonic acid as catalyst), to afford the intermediate glycosyl acceptor, (3), which was coupled with 2,3-di-*O*-acetyl-4,6-*O*-benzylidene-glucopyranosyl trichloroacetimidate, (4) (Larson & Heathcock, 1997), under silver triflate catalysis to generate the desired disaccharide, (II), in 50% yield. Spectroscopic analysis for (II), ${}^1\text{H}$ NMR (CDCl_3 , 300 MHz, δ , p.p.m.): 1.3 (*s*, 3H), 1.4 (*d*, 3H), 1.5 (*s*, 3H), 2.0, 2.1 (*s*, 6H), 3.4 (*m*, 1H), 3.7 (*m*, 4H), 3.9 (*dd*, 1H), 4.0 (*m*, 2H), 4.3 (*d*, 1H), 4.6 (*d*, 1H), 4.9 (*t*, 2H), 5.0 (*t*, 1H), 5.4 (*s*, 1H), 7.3 (*m*, 10H).

Compound (I)

Crystal data

$\text{C}_{17}\text{H}_{22}\text{O}_7$
 $M_r = 338.36$
Monoclinic, $P2_1$
 $a = 8.735$ (2) Å
 $b = 9.526$ (2) Å
 $c = 10.605$ (3) Å
 $\beta = 91.00$ (2)°
 $V = 882.4$ (5) Å³
 $Z = 2$

$D_x = 1.273$ Mg m^{–3}
Mo $K\alpha$ radiation
Cell parameters from 15 reflections
 $\theta = 11.2$ – 12.6 °
 $\mu = 0.10$ mm^{–1}
 $T = 293$ (2) K
Prism, colourless
 $0.85 \times 0.70 \times 0.32$ mm

Data collection

Enraf–Nonius CAD-4 diffractometer
Non-profiled $\omega/2\theta$ scans
Absorption correction: ψ scan (North *et al.*, 1968)
 $T_{\min} = 0.890$, $T_{\max} = 0.965$
2982 measured reflections
2820 independent reflections
2457 reflections with $I > 2\sigma(I)$

$R_{\text{int}} = 0.017$
 $\theta_{\text{max}} = 30.4$ °
 $h = 0 \rightarrow 12$
 $k = -13 \rightarrow 0$
 $l = -15 \rightarrow 15$
3 standard reflections
frequency: 60 min
intensity decay: none

Refinement

Refinement on F^2
 $R(F) = 0.040$
 $wR(F^2) = 0.130$
 $S = 1.09$
 2820 reflections
 221 parameters
 H-atom parameters constrained

$$w = 1/[\sigma^2(F_o^2) + (0.0964P)^2 + 0.0099P]$$

where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\max} = 0.004$
 $\Delta\rho_{\max} = 0.23 \text{ e } \text{\AA}^{-3}$
 $\Delta\rho_{\min} = -0.16 \text{ e } \text{\AA}^{-3}$

Table 1

Selected geometric parameters (\AA , $^\circ$) for (I).

C1—O1	1.394 (2)	C5—O5	1.434 (2)
C1—O5	1.411 (2)		
O1—C1—O5	108.78 (13)	O5—C5—C4	109.90 (14)
O1—C1—C2	106.60 (14)	C1—O1—C11	114.44 (16)
O5—C1—C2	111.65 (12)	C1—O5—C5	111.84 (12)
O5—C1—C2—C3	57.13 (16)	C3—C4—C5—O5	-55.72 (17)
C2—C3—C4—C5	53.06 (17)	C12—C11—O1—C1	-62.4 (3)

Table 2

Hydrogen-bonding geometry (\AA , $^\circ$) for (I).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
O2—H2A \cdots O7 ⁱ	0.82	2.14	2.878 (2)	150
C15—H15 \cdots O5 ⁱⁱ	0.93	2.57	3.447 (3)	158

Symmetry codes: (i) $2 - x, \frac{1}{2} + y, 1 - z$; (ii) $1 + x, y, z$.

Compound (II)

Crystal data

$C_{33}H_{40}O_{12}$
 $M_r = 628.65$
 Orthorhombic, $P2_12_12_1$
 $a = 8.901 (1) \text{\AA}$
 $b = 14.261 (1) \text{\AA}$
 $c = 25.583 (1) \text{\AA}$
 $V = 3247.4 (4) \text{\AA}^3$
 $Z = 4$
 $D_x = 1.286 \text{ Mg m}^{-3}$

Mo $K\alpha$ radiation
 Cell parameters from 25 reflections
 $\theta = 11.9\text{--}20.9^\circ$
 $\mu = 0.10 \text{ mm}^{-1}$
 $T = 293 (2) \text{ K}$
 Prism, colourless
 $0.65 \times 0.50 \times 0.32 \text{ mm}$

Data collection

Enraf-Nonius CAD-4 diffractometer
 Non-profiled $\omega/2\theta$ scans
 Absorption correction: ψ scan (North *et al.*, 1968)
 $T_{\min} = 0.905$, $T_{\max} = 0.965$
 5471 measured reflections
 5471 independent reflections

2832 reflections with $I > 2\sigma(I)$
 $\theta_{\max} = 30.4^\circ$
 $h = -12 \rightarrow 0$
 $k = -20 \rightarrow 0$
 $l = 0 \rightarrow 36$
 3 standard reflections
 frequency: 60 min
 intensity decay: none

Refinement

Refinement on F^2
 $R(F) = 0.041$
 $wR(F^2) = 0.138$
 $S = 0.89$
 5471 reflections
 411 parameters

H-atom parameters constrained
 $w = 1/[\sigma^2(F_o^2) + (0.0882P)^2]$
 where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\max} = 0.022$
 $\Delta\rho_{\max} = 0.21 \text{ e } \text{\AA}^{-3}$
 $\Delta\rho_{\min} = -0.19 \text{ e } \text{\AA}^{-3}$

The absolute configurations of these two light-atom structures were not determined by the present X-ray analyses but were already known from the known configurations of the starting materials. H atoms were placed in calculated positions, with C—H distances in the range 0.93–0.98 \AA and an O—H distance of 0.82 \AA , and their

Table 3

Selected geometric parameters (\AA , $^\circ$) for (II).

C1—O1	1.390 (3)	C17—O10	1.419 (3)
C1—O5	1.418 (3)	C17—C18	1.513 (4)
C2—O2	1.432 (3)	C18—C19	1.536 (3)
C5—O5	1.424 (3)	C19—C20	1.504 (4)
C17—O2	1.396 (3)	C20—C21	1.522 (3)
O1—C1—O5	108.0 (2)	O10—C17—C18	109.1 (2)
O1—C1—C2	109.6 (2)	O10—C21—C22	110.8 (2)
O5—C1—C2	108.0 (2)	C1—O1—C10	111.29 (19)
O5—C5—C4	111.2 (2)	C17—O2—C2	114.62 (18)
O2—C17—O10	108.6 (2)	C1—O5—C5	111.3 (2)
O2—C17—C18	108.3 (2)	C17—O10—C21	109.07 (19)
O5—C1—C2—C3	61.6 (3)	O10—C17—O2—C2	-64.5 (3)
C1—C2—C3—C4	-44.8 (3)	C3—C2—O2—C17	-65.9 (3)
C2—C3—C4—C5	36.2 (3)	C2—C1—O5—C5	-72.1 (3)
C3—C4—C5—O5	-42.4 (3)	C4—C5—O5—C1	61.2 (3)
C17—C18—C19—C20	-48.9 (3)	C18—C17—O10—C21	-65.6 (3)
C18—C19—C20—C21	51.1 (3)	C20—C21—O10—C17	69.5 (3)
C19—C20—C21—O10	-61.8 (3)		

Table 4

Hydrogen-bonding geometry (\AA , $^\circ$) for (II).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
C17—H17 \cdots O3	0.98	2.35	2.995 (3)	123
C22—H22A \cdots O12 ⁱ	0.97	2.59	3.095 (4)	113

Symmetry code: (i) $1 + x, y, z$.

isotropic displacement parameters were fixed at 1.2 or 1.5 times the U_{eq} of the bonded atom.

For both compounds, data collection: *CAD-4 EXPRESS* (Enraf-Nonius, 1994); cell refinement: *CAD-4 EXPRESS*; data reduction: *XCAD4* (Harms & Wocadlo, 1995); program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEP-3 for Windows* (Farrugia, 1997) and *PLATON* (Spek, 1998); software used to prepare material for publication: *WinGX* (Farrugia, 1999).

Supplementary data for this paper are available from the IUCr electronic archives (Reference: FG1655). Services for accessing these data are described at the back of the journal.

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